

# Influence of Complement factor H on Factor XIIIA activation: A preliminary study

**Sneha Gupta<sup>1</sup>**, Christoph Krettler<sup>2</sup>, Christoph Reinhart<sup>2</sup>, Johannes Dodt<sup>3</sup>, Andreas Reuter<sup>3</sup>, Vytautas Ivaskevicius<sup>1</sup>, Johannes Oldenburg<sup>1</sup>, Arijit Biswas<sup>1</sup> <sup>1</sup>Institute of Experimental Haematology and Transfusion Medicine, University Clinic Bonn, Germany. <sup>3</sup>Paul Ehrlich Institute, 63225 Langen, Germany.

# Introduction and Objective

Fibrogammin P, the plasma concentrate of Coagulation Factor XIII (FXIII), is used for treating FXIII deficient patients. Biochemical content characterization of Fibrogammin P has shown Complement factor H (CFH) to be one of its major constituents. Since CFH is a structurally homologous molecule to FXIIIB, the natural partner of FXIIIA, we contemplate that CFH might influence the functional profile of FXIII. We performed mixing studies, in order to test this hypothesis.

### **Materials and Methods**

In order to purify FXIIIA<sub>2</sub>B<sub>2</sub>, series of **Size exclusion chromatography** was performed till single, monodispersed homogenous peak of the FXIIIA<sub>2</sub>B<sub>2</sub> heterotetramer was obtained. The peak fractions were ran onto SDS PAGE, and bands were confirmed by gel tryptic digestion followed by mass spectrometry (**Figure 1**).

FXIIIAa generation<sup>1</sup> monitored for different plasmatic combinations (mixing studies). To analyse the effect of presence/absence of CFH on FXIII generation in Standard vs. FXIII deficient plasma **rFXIIIA**: Purified from *Pichia Pink*. **rFXIIIB**: Commercially available (Zedira) **Complement Factor H**: Isolated from Fibrogammin P (Figure 1) The resulting activation curves (Figure 2) were evaluated based on a biexponential, mathematical model with first order absorption and elimination. Furthermore, the data was fitted to the equation:  $C(t) = c^*k_a / (K_a - k_b)^* \{[exp($ k<sub>b</sub>\*(t-t<sub>lag</sub>)]- [exp(-k<sub>a</sub>\*(t-t<sub>lag</sub>)]}, where, k<sub>a</sub> is constant of absorption (activation) which describes the development of active FXIIIAa species, and  $k_{\rm b}$  is the elimination (deactivation) constant (Figure 3).

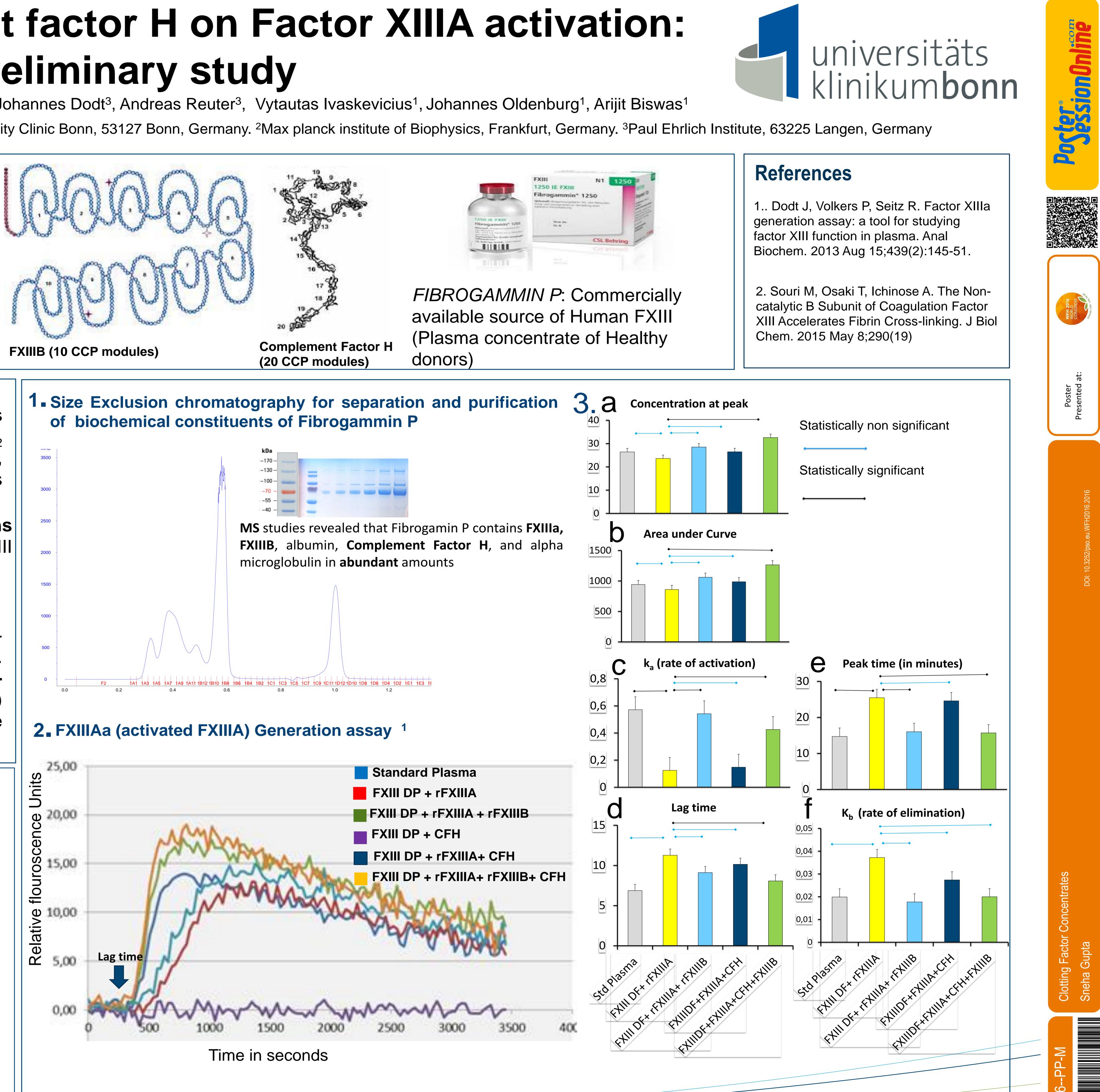
## Results

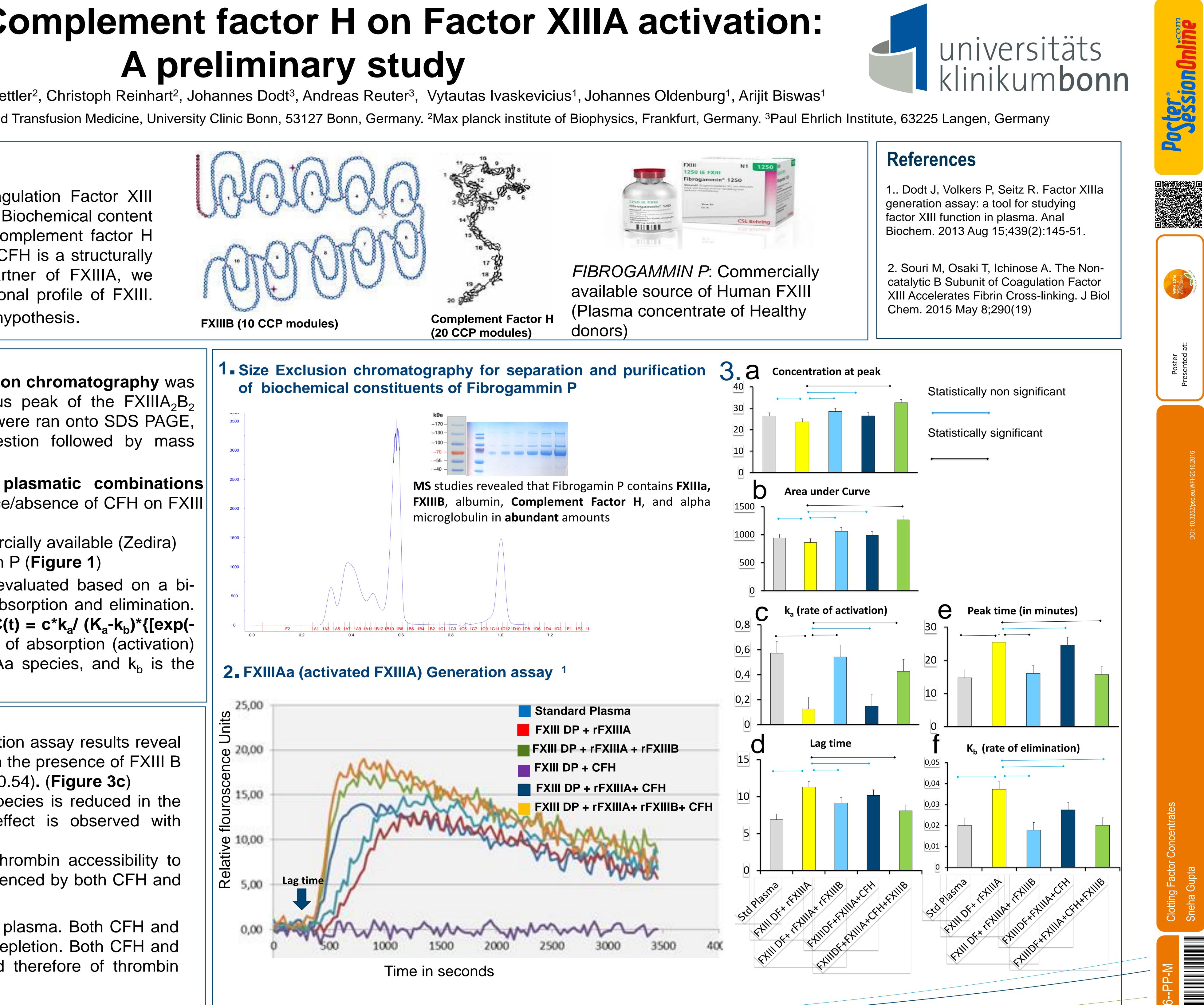
- . In agreement with an earlier report<sup>2</sup> the generation assay results reveal that, rate of activation of FXIIIA is accelerated in the presence of FXIII B
- (Ka (FXIIIDP+ FXIIIA) is 0.12, Ka (FXIIIDP+ FXIIIA+FXIIIB) is 0.54). (Figure 3c)
- 2. However, the rate of depleteion of activated species is reduced in the presence of FXIIIB subunit and a similar effect is observed with CFH(**Figure 3f**).
- 3. The lag time (Figure 3d), which represents thrombin accessibility to FXIIIA molecule in the assay is also mildly influenced by both CFH and FXIIIB.

#### Conclusion

FXIIIB accelerates the rate of FXIIIA activation in plasma. Both CFH and

FXIIIB appears to influence the rate of FXIIIAa depletion. Both CFH and FXIIIB additively control access to thrombin and therefore of thrombin mediated cleavage of FXIIIA.





Authors acknowledge funding from CSL Behring for reasearch on Factor XIII