Haemoglobin induces podocyte injury in mice and humans with pathologies associated with massive intravascular hemolysis.

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Background

Recurrent and massive intravascular hemolysis is associated with progressive impairment of renal function, proteinuria and glomerulosclerosis, suggesting podocyte involvement. Podocytes are highly differentiated epithelial cells that play a key role in preserving glomerular filtration barrier. Structural alterations in podocyte foot processes occur in many glomerular diseases.

However, the cellular and molecular mechanisms involved in Hb-renal damage remains unclear. Control of ROS is important to restore the balance between oxidants and antioxidants in cells. In this regard, nuclear factor erythroid-2-related factor 2 (Nrf2) plays a central role in the defense against oxidative stress by activating the expression of a number antioxidants and phase 2 detoxifying enzymes.

Intravascular hemolysis promoted Hb accumulation and heme catabolism activation in podocytes *in vivo*







Aims

Our objective was to determine the capacity for Hb uptake by murine podocyte *in vivo* and *ex vivo*, its injurious actions and the molecular pathways involved in this context. We also analyzed whether Hb-loaded podocytes are detected in kidney biopsies and urinary sediments from patients with severe intravascular hemolysis.





Hemolysis was induced in 12-week-old C57BL/6 and Nrf2 knockout mice by intraperitoneal administration of phenylhydrazine (200mg/kg of body weight, Sigma-Aldrich). Mice were euthanized 24 hours after phenylhydrazine injection (n=5 per group) and blood and urine samples were collected for biochemistry; and dissected kidneys for RNA and protein expression, flow cytometry analysis, immunofluorescence, and transmission electronic microscopy (A) Representative picture of the kidneys and the experimental model scheme. (B) Biochemical characteristics of Nrf2+/+ and Nrf2-/- mice treated or not with phenylhydrazine. (C) Representative confocal microscopy images showing co-localization (white arrows) of Hb (green), HO-1 (green), Ferritin (green) with the podocyte marker nephrin (red) in Phe-mice, scale bar 25 µM. Nuclei were stained with DAPI (blue). (D) Representative images of transmission electronic microscopy showing fusion of podocyte foot processes in phenylhydrazine-treated mice (Phe). Results are expressed as mean ± SE. * p<0.05 vs control mice. Animal studies were in accordance with the Directive 2010/63/EU of the European Parliament and were approved by the Institutional Animal Care and Use Committee (IIS-Fundacion Jimenez Diaz)

Nrf2 knockout mice have increased podocyte death and altered expression of key proteins for podocyte function





Murine Podocytes bound and endocytosed Hb in an energy- and active receptor-dependent way





(A) Representative immunofluorescence images showing co-localization of M30 (green), a marker of apoptosis, with the podocyte marker nephrin (red) in saline or Phenylhydrazine (Phe)-treated mice (n=5 per group). Scale bar 25 µM. (B) Semi-quantification of M30 staining in glomerular cross sections. (C) Representative immunofluorescence of Nephrin (red), WT-1 (green), and DAPI (blue) in glomeruli from both control- and Phe-mice, scale bar 25 µM. (D) Quantitative analysis of podocyte number (WT-1 positive cells) in glomeruli from hemolytic-mice. (E-F) Synaptopodin (Syn) and Nephrin (Neph) protein and mRNA expression measured by western blot and RT-qPCR, respectively. Results (mean ± SE) are expressed as fold time increase as compared to control mice.

Podocytes were stained for Hb, pNrf2 and HO-1 in renal sections of patients with massive intravascular hemolysis



Mice podocytes were isolated from kidneys by a size exclusion protocol, and cultured in serum-free media and incubated with different concentrations of podocytes by confocal microscopy. Synaptopodin (green) and DAPI (blue) were used to determine podocyte cells and nuclei, respectively. Data are expressed as mean±standard deviation. *p<0,05 vs Control, \neq p<0,05 vs Hemoglobin 50µg/mL)

Podocytes were stained for Hb in urinary sediments of patients with massive intravascular hemolysis





Representative confocal microscopy images showing co-localization (white arrows) of Hb (red) (A), pNrf2 (B) and HO-1 (red) (C) with the podocyte marker synaptopodin (green) in patients with aHUS and healthy controls, scale bars 50 µM. Nuclei were stained with DAPI (blue). The rectangle shows the region of interest



(A) Urine sediments from both healthy individuals and aHUS patients were centrifuged onto polylysine-coated slides using a cytospin and immunofluorescence studies were performed. The left panel shows the presence of podocytes (nucleated cells stained with synaptopodin, green, white arrows). High-magnification images are shown in the right panels where Hb (red) is located inside podocytes (synaptopodin, green) from urine sediments of aHUS patients, as determined by confocal microscopy. DAPI (blue) was used to stain nuclei. Scale bars 10µm. (B) Representative confocal microscopy images of urine sediments from patients with paroxysmal nocturnal hemoglobinuria showing the presence of Hb (red) inside podocytes (nucleated cells stained with synaptopodin, green), scale bars 10 μ M.

Conclusion

In conclusion, our study identifies podocytes as new targets of intravascular hemolysis. Moreover, Nrf2 activation may be a potential therapeutic target to prevent loss of renal function in patients with severe intravascular hemolytic crisis. These findings provide new insights into novel aspects of Hb-toxicity and may have important pathogenic and therapeutic implications for intravascular hemolysis related diseases.

