

Discrepancy of F8 and F9 gene variant classifications between clinical laboratories

Introduction and Objectives:

Despite the F8 and F9 genes being well-conserved, not all variants in these genes are pathogenic. Targeted, familial genetic testing for variants which are not pathogenic may lead to inaccurate risk assessment and genetic counseling for relatives. Therefore, variant classification is a critical element for determining a genetic test result's clinical utility.

There are multiple professional guidelines and standards that exist for clinical genetic test results, including recommendation to classify an identified sequence variant according to a 5-tier system (Figure 1) and, more recently, a suggested classification process utilizing various types of supporting evidence.¹⁻³ However, discrepancy of variant classification between laboratories has been demonstrated in other genetic diseases.⁴

We review the experience of our Hemophilia Treatment Center (HTC) with patients who have discrepant F8 and F9 variant classifications between clinical laboratories, as well as our approaches for addressing this potential clinical dilemma.



Figure 1. Five-tier system for classifications of sequence variants ^{2,3}

Methods and Materials:

Patients with hemophilia A or B from our HTC who had a genetic test result, including a variant classification, from more than one clinical laboratory were identified. Individuals identified to have one of the following well-defined F8 and F9 variants were excluded: F8 intron 22 inversion, F8 intron 1 inversion, F9 Amish founder mutation.⁵⁻⁷ Classification and supporting lines of evidence for test results were compared for each individual.

ML Alabek¹, MV Ragni^{1,2}, LM Malec^{1,2}, C Seaman^{1,2}

Hemophilia Center of Western Pennsylvania, Pittsburgh PA¹, University of Pittsburgh, Pittsburgh PA²

Results:

Twelve patients were identified with ten unique variants. Two variants were each identified in two patients, not known to our HTC to be related. Eligible test results were from 3 different laboratories.

Comparisons of variant classifications are detailed in Table 1.

- Classifications were discrepant for seven of twelve individuals (58.3%).
- All discrepant variants were missense mutations.

Differing lines of evidence between laboratories are outlined in Table 2. Eleven individuals (91.7%) had at least one different line of evidence documented between their two laboratory reports.

DISCREPANT VARIANT CLASSIFICATIONS	F8 (n=2)	F9 (n=5)	Total (n=7)
Classifications Uncertain significance, pathogenic Likely pathogenic, pathogenic	1 1	3* 2^	4* 3^
Mutation type Missense	2	5*^	7*^
Severity Severe Moderate Mild	1 0 1	0 4*^ 1	1 4*^ 2
CONSISTENT VARIANT CLASSIFICATIONS	F8 (n=2)	F9 (n=3)	Total (n=5)
Classification Pathogenic Uncertain significance	2 0	2 1	4 1
Mutation type Missense Nonsense Frameshift Non-coding	1 0 1 0	0 1 1 1	1 1 2 1
Severity Severe Moderate Mild	0 1 1	3 0 0	3 1 1

Table 1. Summary of discrepant and consistent variant classifications among individuals with clinical genetic testing through more than one laboratory. *,^ Denotes that two individuals were identified to have the same variant.

1Dpopulation data, computational and predictive datan/a2^D1n/apopulation data, computational and predictive data, computational and predictive data, segregation data3^D11population data3^D11110111	dictive
2^Dn/apopulation data, computational and predata, segregation data3*Dpopulation datafunctional data	dictive
3 ^{*D} population data functional data	
4 ^c population data, functional data n/a	
5 ^{^D} n/a population data, computational and pre data, segregation data	dictive
6 ^c population data, computational and predictive n/a data	
7 ^D n/a population data, computational and pre	dictive
8 ^D population data, computational and predictive n/a data, segregation data	
9 ^{*D} n/a population data	
10 ^c population data, computational and predictive n/a data	
11 ^C population data computational and predictive data, segr	egation
12 ^c n/a n/a	

Table 2. Summary of lines of evidence for variant classification differing between laboratories. Lab A/B randomly assigned for each case. *, ^ cases identified to have the same variant. ^C, ^D cases with consistent and discrepant classification between laboratories, respectively.

Conclusions:

Variant classification, which may impact a test result's clinical utility, can be laboratory-dependent. To remedy this potential clinical dilemma, our HTC has found it helpful to understand a laboratory's internal processes for variant classification, as well to collaborate on a case-by-case basis with personnel from the laboratory of record. Existing guidelines may help to improve consistency of variant classifications across laboratories; however, it will remain critical for ordering providers to understand variant classification concepts in order to ensure correct diagnosis and interpretation of test results for patients and at-risk family members.

References/Bibliography:

1. Rehm HL, Bale SJ, Bayrak-Toydemir P, et al.; Working Group of the American College of Medical Genetics and Genomics Laboratory Quality Assurance Commitee. ACMG clinical laboratory standards for next-generation sequencing. Genet Med 2013;15:733–747. 2. Richards, S., Aziz, N., Bale, S., Bick, D., Das, S., Gastier-Foster, J., Rehm, H. L. (2015). Standards and guidelines for the interpretation of sequence variants: A joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genetics in Medicine Genet *Med*, 17(5), 405-423. **3.** Richards CS, Bale S, Bellissimo DB, et al.; Molecular Subcommittee of the ACMG Laboratory Quality Assurance Committee. ACMG recommendations for standards for interpretation and reporting of sequence variations: Revisions 2007. Genet Med 2008;10:294–300. 4. Pepin, M. G., Murray, M. L., Bailey, S., Leistritz-Kessler, D., Schwarze, U., & Byers, P. H. (2015). The challenge of comprehensive and consistent sequence variant interpretation between clinical laboratories. Genetics in Medicine Genet Med, 18(1), 20-24. 5. Lakich, D., Kazazian, H. H., Antonarakis, S. E., & Gitschier, J. (1993). Inversions disrupting the factor VIII gene are a common cause of severe haemophilia A. Nature Genetics Nat Genet, 5(3), 236-241. 6. Bagnall, R. D. (2002). Recurrent inversion breaking intron 1 of the factor VIII gene is a frequent cause of severe hemophilia A. Blood, 99(1), 168-174. 7. Ketterling, R., Bottema, C., Koeberl, D., Ii, S., & Sommer, S. (1991). T296----M, a common mutation causing mild hemophilia B in the Amish and others: Founder effect, variability in factor IX activity assays, and rapid carrier detection. *Hum Genet Human Genetics*, 87(3).



HOISS

