

Diagnosis of autosomal dominant polycystic kidney disease using efficient PKD1 and PKD2 targeted next generation sequencing

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INTRODUCTION

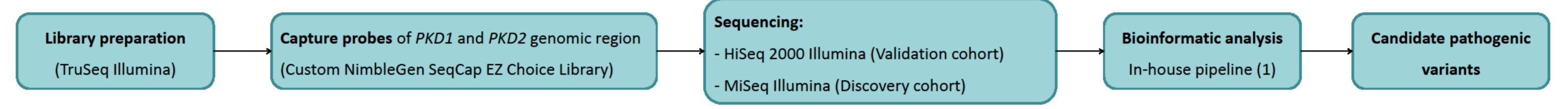
Autosomal polycystic kidney disease (ADPKD) is caused by mutations in PKD1 in 85% of cases and PKD2 in the remaining 15%. Approximately 70% of genomic PKD1 region (exons 1-33) is duplicated 6 times within 6 pseudogens. This, together with the high GC content, the absence of mutation hot spots and the high allelic heterogeneity, makes the molecular diagnostics of ADPKD challenging. To date, sequencing of PKD1 repeated region requires long-range amplifications followed by nested PCRs, combined with Sanger sequencing of all 46 PKD1 and 15 PKD2 exons. If no mutations are identified, multiplex ligation-dependent probe amplification (MLPA) analysis is performed to identify potential insertions and deletions.

AIMS

- To validate NGS technology for genetic diagnostics of ADPKD in a validation cohort: 36 patients with known mutations in PKD1 and PKD2
- To apply this approach in a discovery cohort including 12 ADPKD patients with unknown mutations

MATERIALS AND METHODS

Assay workflow



Pathogenicity evaluation

All the variants were validated by Sanger sequencing

- Mutations that could give rise to premature protein (truncating mutations)
- Missense and noncanonical splicing variants → SCORING SYSTEM →
- DEFINITELY PATHOGENIC
 - HIGHLY LIKELY PATHOGENIC
 - LIKELY PATHOGENIC
 - INDETERMINATE VARIANTS - NEUTRAL VARIANTS

RESULTS

Panel design

- The capture probes designed covered all the coding regions
- Capture probes could not be designed in 19% of intronic

Depth of coverage quences

	Mean (x)	≥100x (%)	≥50x (%)	≥20x (%)	
VALIDATION COHORT					
PKD1	331	85	92	96	
PKD2	481	97	99	99	
DISCOVERY COHORT					
PKD1	81	52	65	87	
PKD2	174	68	93	99	

Detection of PKD1 and PKD2 mutations

Conservation in orthologs

- ADPKD mutation database

- In silico predictors: SIFT, Polyphen

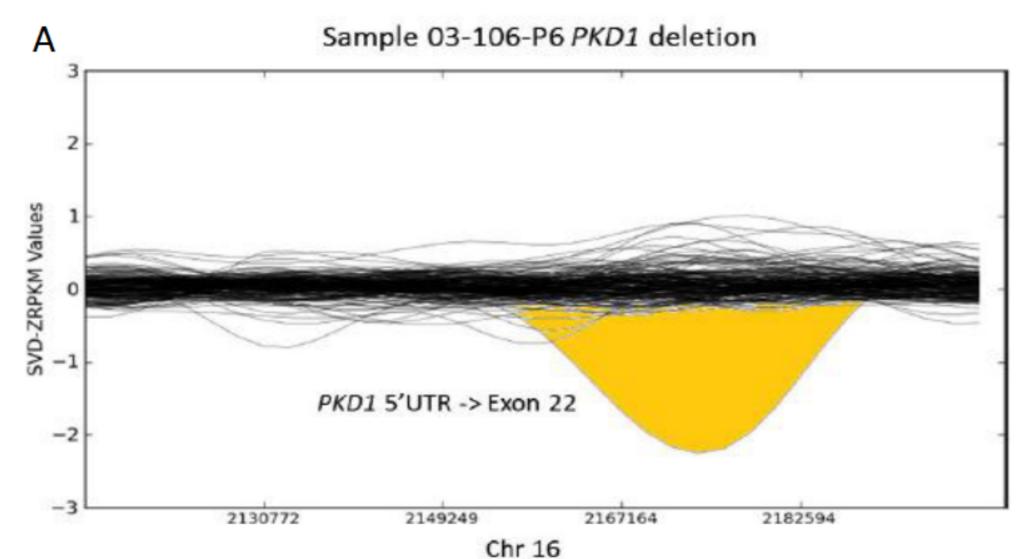
- Allele frequency: dbSNP, 1000 genomes

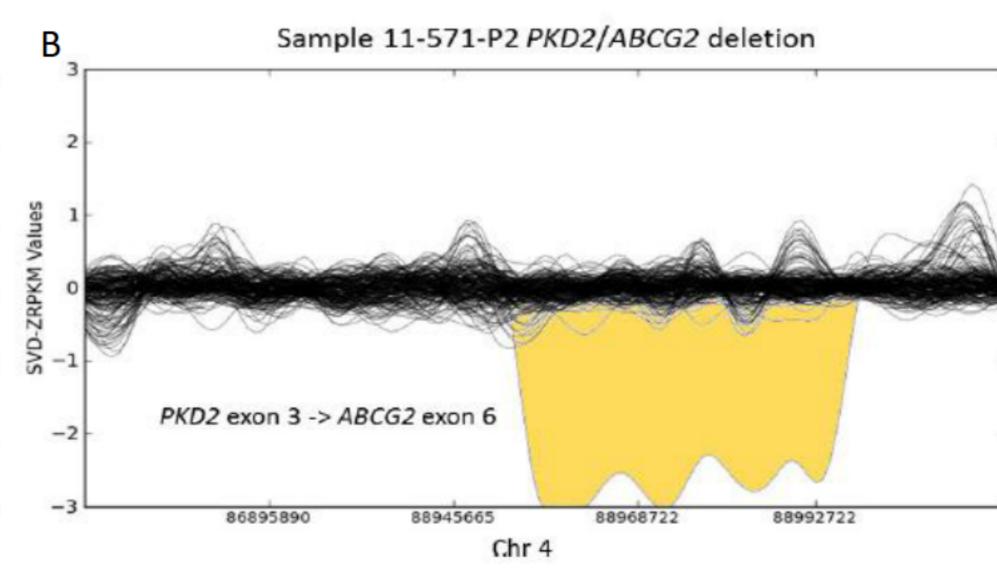
	Detected	Not detected
VALIDATION COHORT		
Nº of mutations in <i>PKD1</i> duplicated region	24	1
Nº of mutations in <i>PKD1</i> no duplicated region	6	0
Nº of mutations in <i>PKD2</i>	5	0
Total (%)	35 (97)	1 (3)
DISCOVERY COHORT		
Nº of mutations in <i>PKD1</i> duplicated region	8	1
Nº of mutations in <i>PKD1</i> no duplicated region	2	1
Nº of mutations in <i>PKD2</i>	1	0
Total (%)	11(85)	2 (15)

- Validation cohort: 35 out of 36 previously known different mutations were identified in their correct heterozygous state.
- Discovery cohort: 11 out of 12 mutations were detected in 12 patients (1 patient carried a mutation in *PKD1* and another in *PKD2*).
- Not detected mutations: the 3 mutations not detected by our NGS approach were located in poorly covered regions: exon 1, 23 and 44 of PKD1

Detection of large deletions

Two previously known large deletions were correctly identified (yellow peaks). The normalized relative copy number of an exon in a sample (SVD-ZRPKM values) for the 36 patients of the validation cohort is shown. (A) Patient 03-106-P6 PKD1 deletion (g.2154344-2186386). (B) Patient 11-571-P2: PKD2/ABCG2 deletion (g.88952828-89050618)





CONCLUSIONS

- In-solution capture coupled to NGS is an efficient approach to detect all types of ADPKD pathogenic mutations, including structural variants.
- Our approach is cost- and time- effective, and meets the sensitivity and specificity criteria required for genetic diagnostics of ADPKD.

References

(1) Trujillano D et al 2013 J Med Genet 50(7):455-62

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