

Highly sensitive, selective and rapid LC-MS method for simultaneous quantification of diadenosine polyphosphates in human plasma

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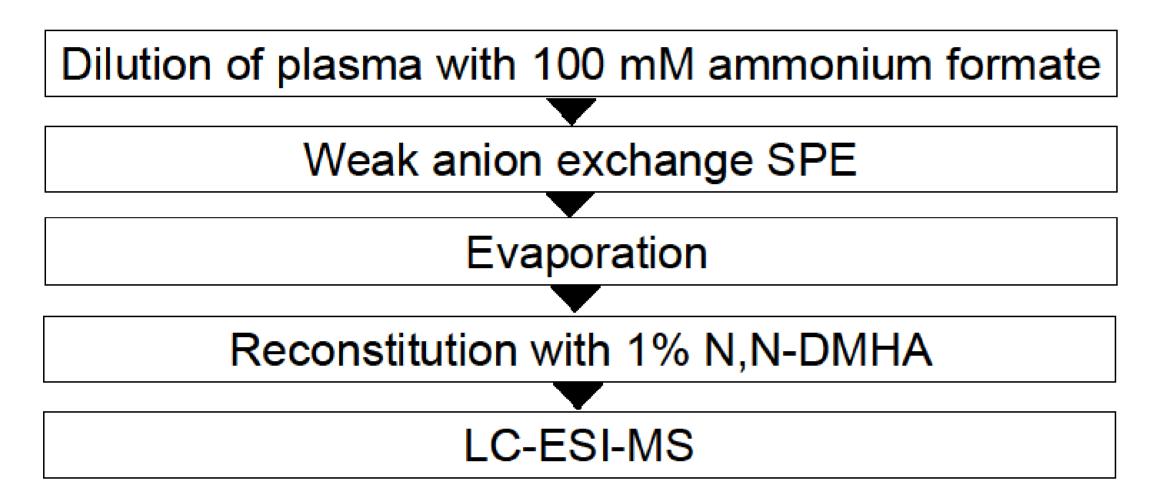


BACKGROUND

Diadenosine polyphosphates (ApnAs) are endogenous mediators involved in a large number of pathophysiologic processes^[1,2]. In plasma, ApnAs are quantified using chromatographic and UV based methods with low selectivity due to co-eluting plasma components and requires high volume. Therefore, we developed and validated a highly sensitive, selective and rapid LC–ESI–MS method for simultaneous quantification of ApnAs (withn = 3–6) in human plasma by comparison of ESI-MS/MS fragment spectra of isolated endogenous compounds with those of pure ApnAs.

METHODS

Diadenosine polyphosphates were extracted from 100µl human plasma using weak anion exchange extraction cartridges. The separation of ApnAs was achieved using capillary C18 columns. ESI-HCT mass spectrometer (Bruker Daltonik, Germany) operated in negative ion mode was used for detection and quantification of ApnAs.



Steps involved in preparation of plasma samples for the quantification of diadenosine polyphosphates

RESULTS

The chemical structure of ApnAs is given in Figure 1. A calibration curve was established for diadenosine polyphosphate free plasma in the concentration range of 1.9-125 nM (r2> 0.998) for all analytes (figure 4). The quantitative determination of the matrix effects revealed minor influence was quantified as -1.1 ± 2.6 for Ap3A, 0.9 ± 2.4 for Ap4A, -3.2 ± 1.7 for Ap5A and 0.3 ± 2.7 for Ap6A (% \pm SEM each) (Figure 3). The intra- and inter-day accuracies were in the range of 91.4% and 110.9%. The intra- and inter-day precisions were determined as 0.1% and 11.4%, respectively (Table 1). The mean plasma concentrations of ApnAs were quantified as 31.9 ± 5.9 nM for Ap3A, 40.4 ± 6.6 nM forAp4A, 10.7 ± 1.5 nM for Ap5A and 10.0 ± 18.9 nM for Ap6A.

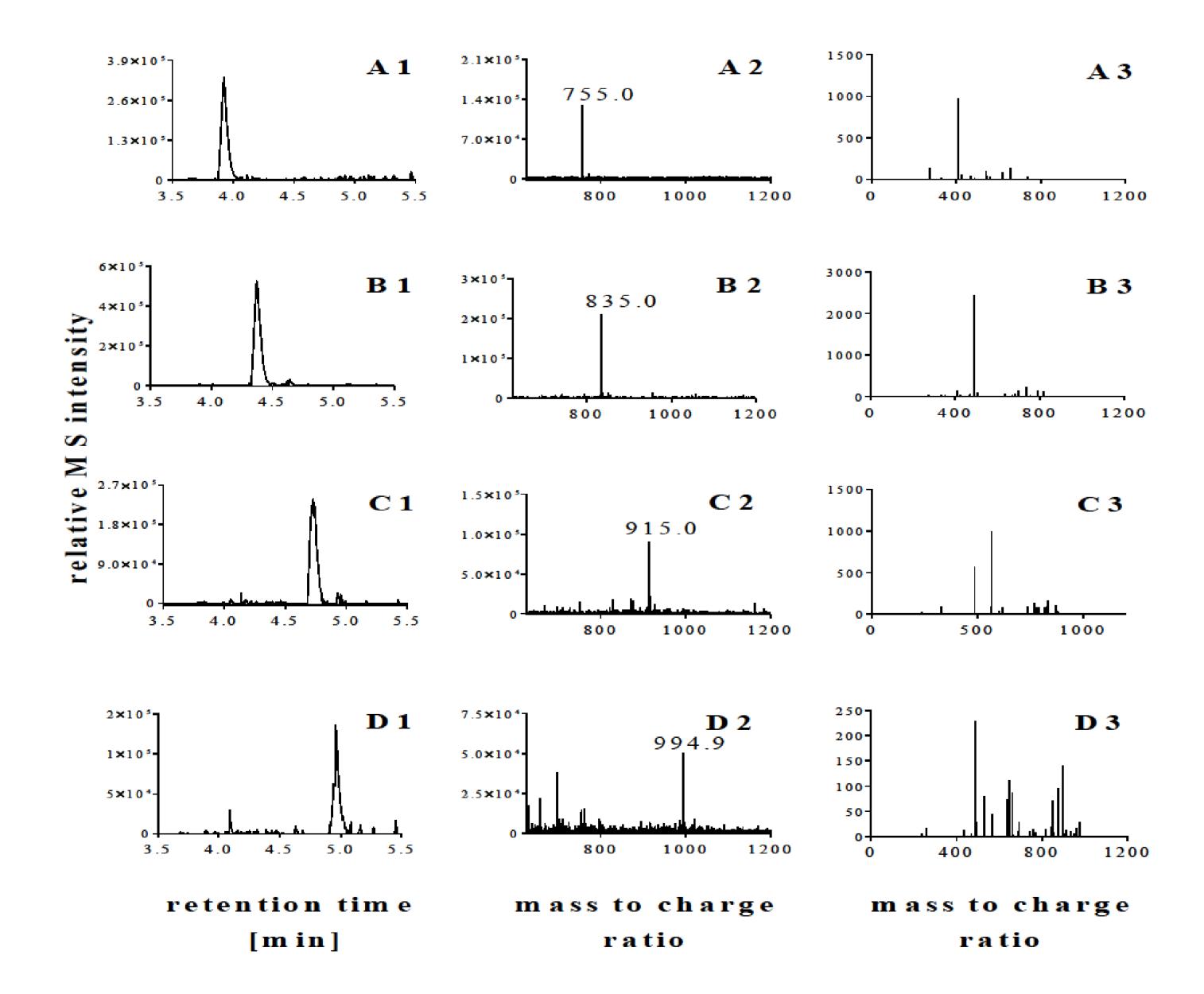


Figure. 2: Representative extracted ion chromatograms of endogenous diadenosine polyphosphates (A1, B1, C1, D1) as well as the corresponding negative ion ESI-MS (A2, B2,C2, D2) and ESI-MS/MS spectra (A3, B3, C3, D3).

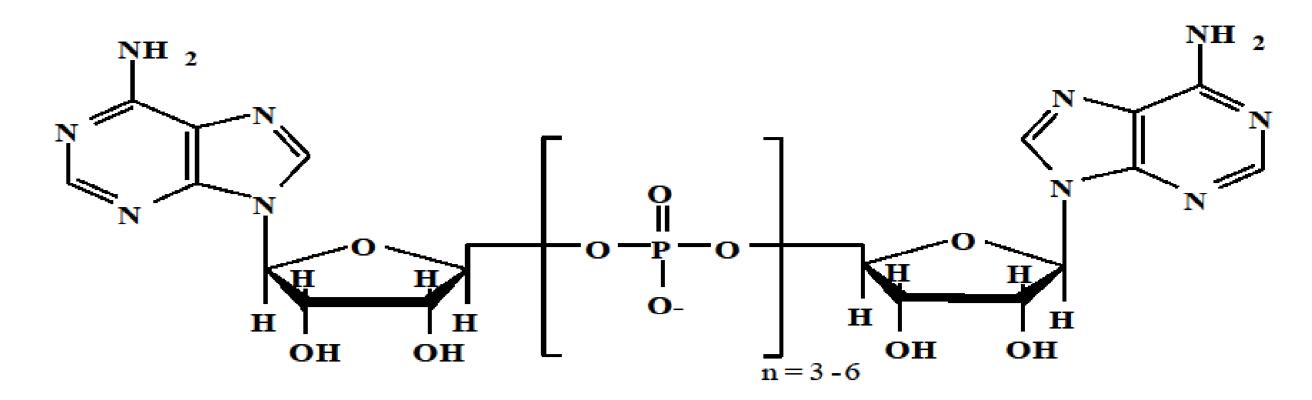


Figure 1: Molecular structure of diadenosine polyphosphates (Ap3–6A)

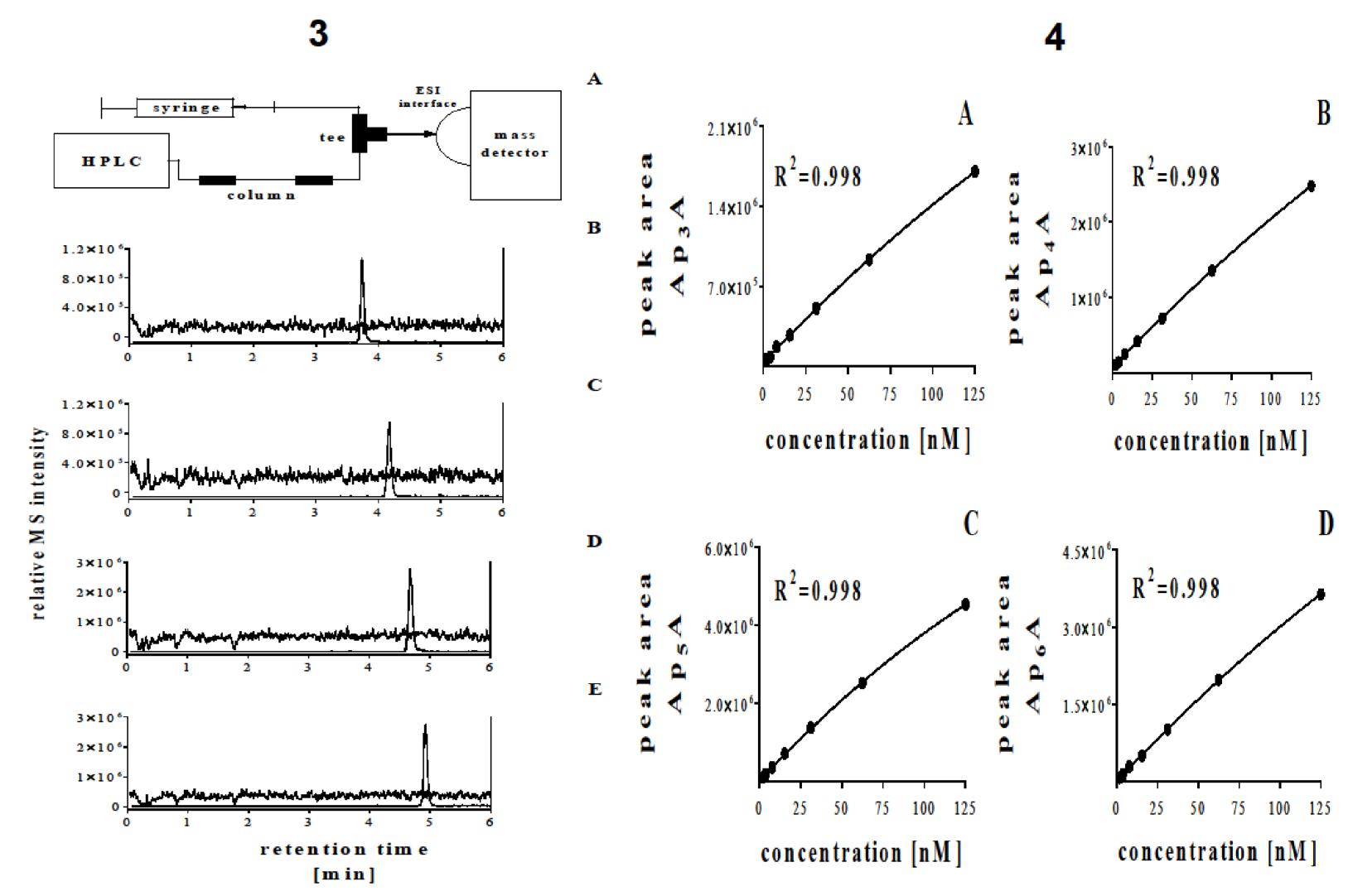


Figure 3: Investigation of ion suppression effects by the post column infusion method. (A) Scheme of the post-column infusion system; (B)–(E) overlay chromatograms showing the matrix effects due to endogenous plasma compounds for Ap3A (B), Ap4A (C), Ap5A (D) and Ap6A (D). Figure 4: Calibration curves of Ap3A (A), Ap4A (B), Ap5A (C) and Ap6A (D)

Table 1: Accuracy and precision of quality control samples

			Intra-day (n=3)		Intrer-day (n=9)	
Analyte	QC	concentration	accuracy	precision	accuracy	precision
		(nM)	(%) ¹	(%) ²	(%) ¹	(%) ²
Ap_3A	LLOQ	1.9	109.6	5.1	106.6	9.6
	LQC	12.5	95.5	6.2	96.2	6.2
	MQC	50	100.3	3.8	102.0	4.0
	HQC	100	109.7	1.0	106.1	3.4
Ap_4A	LLOQ	1.9	108.2	1.7	102.4	10.5
	LQC	12.5	100.0	8.6	97.7	7.3
	MQC	50	101.9	2.5	102.9	1.6
	HQC	100	106.9	0.6	105.2	0.9
Ap ₅ A	LLOQ	1.9	100.6	11.4	105.4	11.4
	LQC	12.5	100.9	5.1	97.1	3.49
	MQC	50	97.7	3.9	97.8	3.2
	HQC	100	110.9	1.3	106.2	0.9
$\mathbf{Ap_6A}$	LLOQ	1.9	106.3	6.4	108.9	8.7
	LQC	12.5	90.5	1.62	91.4	3.9
	MQC	50	107.0	2.42	101.0	3.2
	HQC	100	91.8	0.1	98.8	0.8

¹calculated as (mean determined amount/nominal amount×100)

CONCLUSIONS

Since this method requires low volume of plasma to quantify the diadenosine polyphosphates, highly applicable for clinical applications and also for basic science to study its impact during pathophysiological conditions.

REFERENCES

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- 2. V. Jankowski, M. van der Giet, H. Mischak, M. Morgan, W. Zidek, J. Jankowski, Br. J. Pharmacol. 157 (2009) 1142.

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Disclosure Statement of Financial Interest: I, The authors have a financial interest or affiliation with one or more organizations that could be perceived as a real or apparent conflict of interest in the context of the subject of this presentation









²calculated as % CV. (SD./mean)×100