

Alexander Lukasz<sup>1,2</sup>, Carina Hillgruber<sup>3</sup>, Hans Oberleithner<sup>2</sup>, Kristina Kusche-Vihrog<sup>2</sup>,  
Hermann Pavenstädt<sup>1</sup>, Tobias Görge<sup>3</sup>, Philipp Kümpers<sup>1</sup>

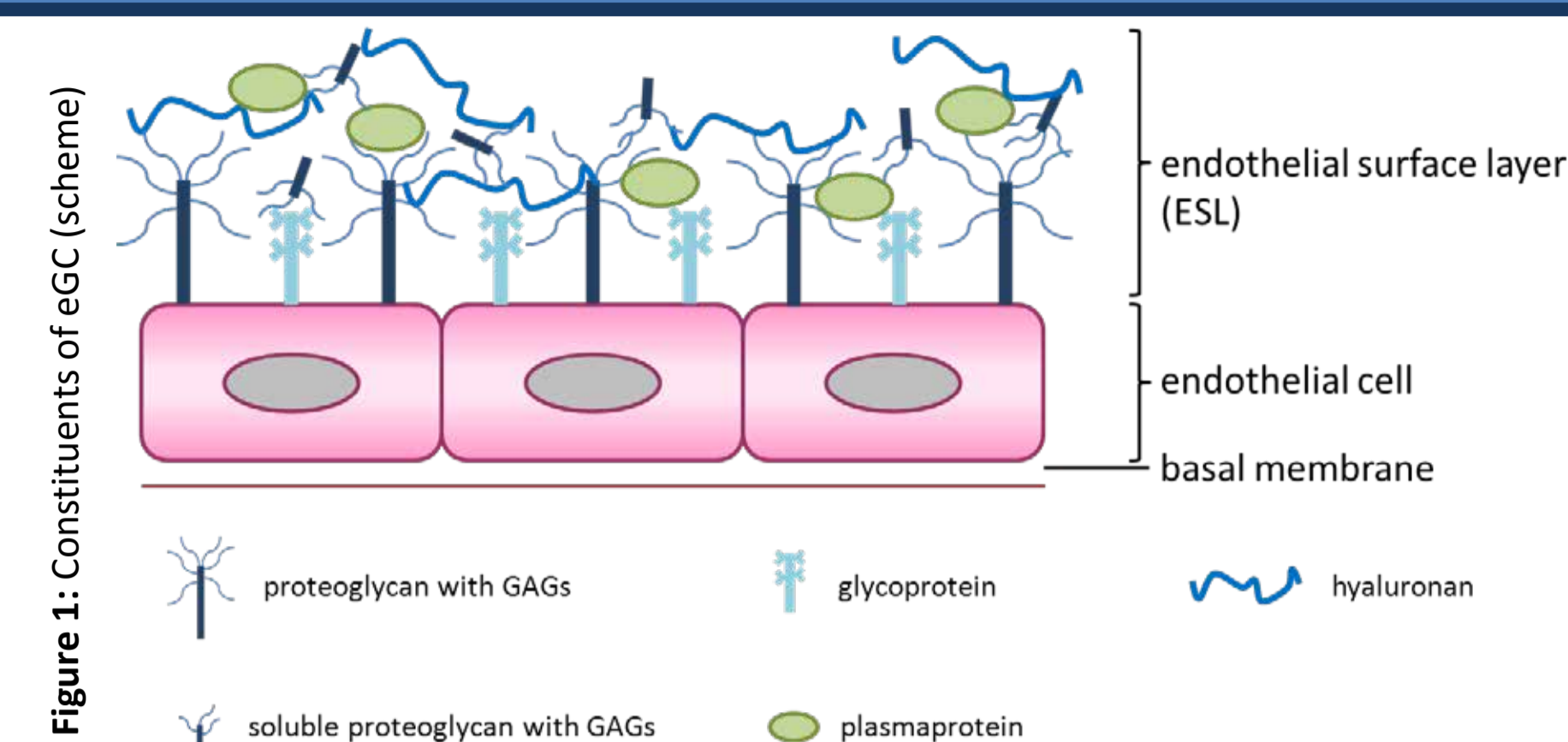
<sup>1</sup> Medizinische Klinik D, Abteilung für Allg. Innere Medizin, Nephrologie und Rheumatologie, Universitätsklinikum Münster, Münster, Deutschland

<sup>2</sup> Institute of Physiology II, University Hospital Münster, Robert-Koch-Straße 27b, 48149 Münster, Germany

<sup>3</sup> Department of Dermatology, University Hospital Münster, Von-Esmarch-Straße 58, 48149 Münster, Germany

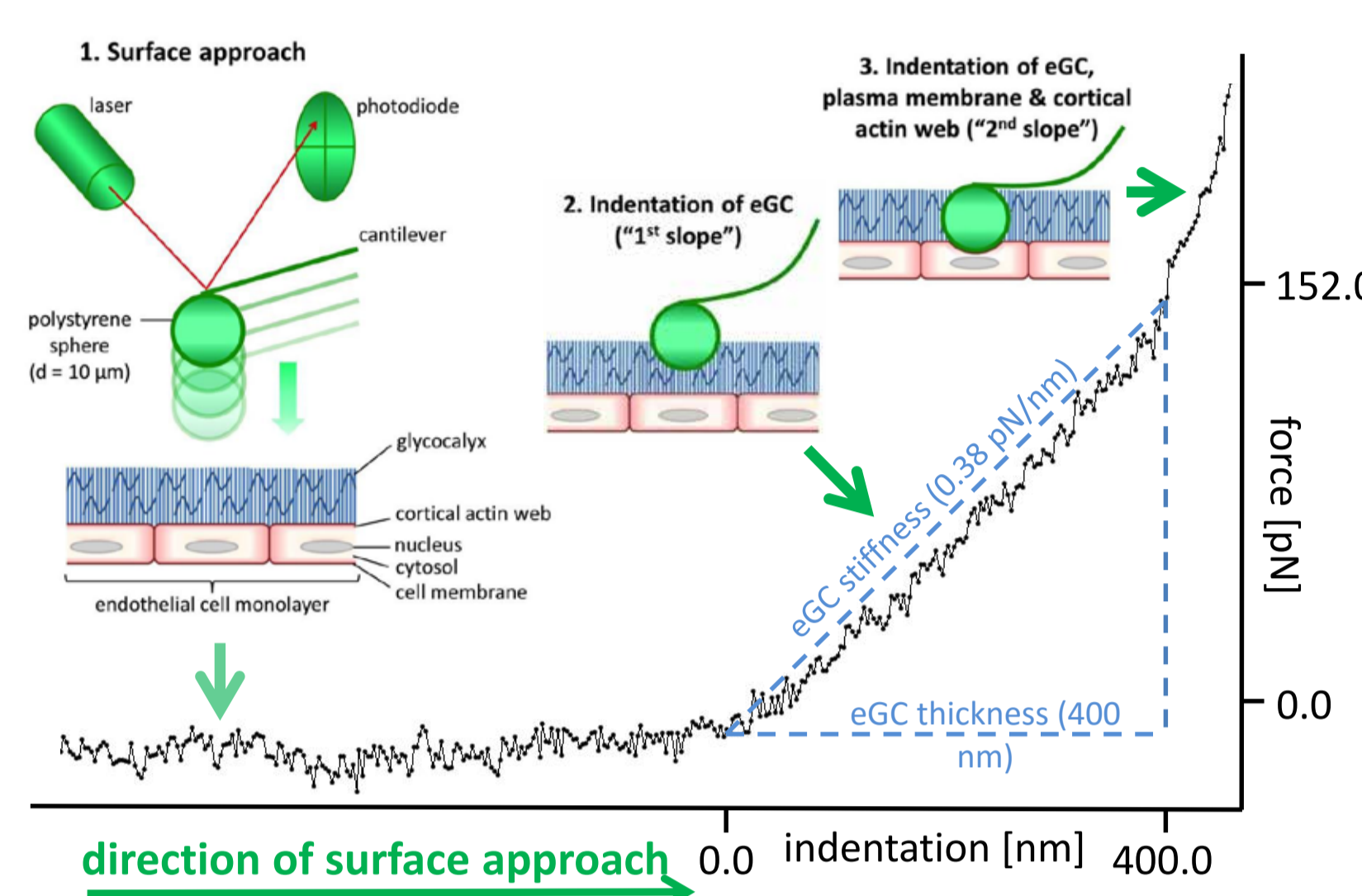
## Introduction

Angiotensin-2 (Angpt-2), an antagonist of the endothelium-stabilizing receptor Tie2 secreted by endothelial cells, promotes vascular permeability. We have previously shown that administration of Tie2 agonistic molecules, which counteract the devastating effects of Angpt-2, prevent ischemic and septic acute kidney injury (AKI). Angpt-2 probably mediates AKI through endothelial-cell contraction and junctional disintegration. Here we hypothesized that Angpt-2 might mediate the breakdown of the endothelial glyocalyx (eGC), a carbohydrate-rich vasoprotective layer lining the luminal surface of the endothelium, as well.



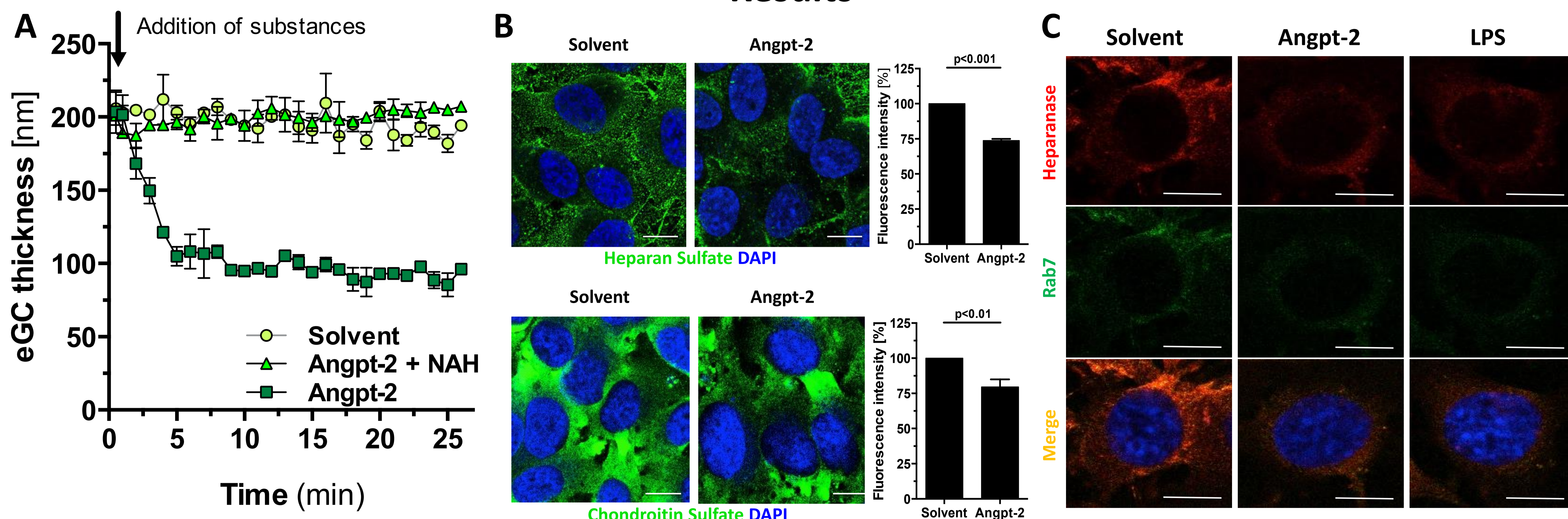
## Methods

- Confocal and atomic force microscopy (figure 2) were used to visualize and analyze the thickness of the eGC on living endothelial cells.
- An *in vivo* permeability assay was used to quantify vascular permeability in murine back skins (miles assay).



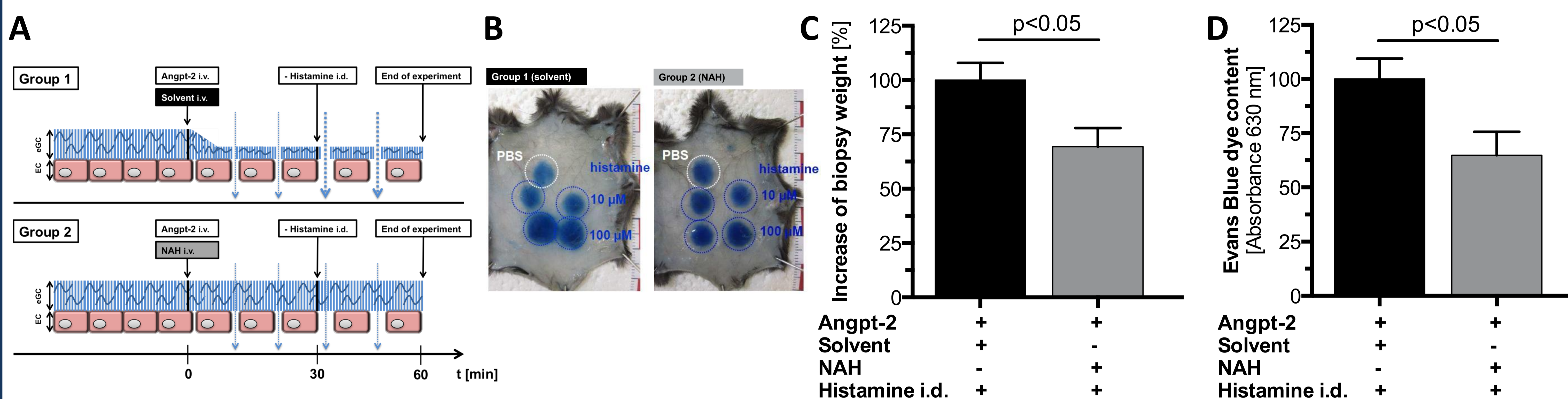
**Figure 2:** Original tracing of a force-distance curve performed on the endothelial surface of living endothelial cells *in vitro*. Using a mechanical nanosensor (cantilever), mounted on an atomic force microscope, height (nm) and stiffness (pN/nm) of the eGC on the luminal surface of endothelial cells were quantified.

## Results



**Figure 3:** Angpt-2 mediates eGC breakdown *in vitro*. In AFM online experiments Angpt-2 alone induces a rapid loss of the eGC in endothelial cells *in vitro* in comparison to Solvent or Angpt-2 and NAH, a substrate of the eGC digesting enzyme heparanase (A). Glycocalyx deterioration involved the specific loss of its main constituents heparan sulfate and chondroitin sulfate (B) paralleled by the secretion of the heparan sulfate-specific glucuronidase heparanase from late endosomal/lysosomal stores (C).

## In-vivo miles assay – edema formation and plasma leakage



**Figure 4:** The principle of the *in vivo* miles assay is schematically shown in (A). Representative pictures of the murine back skin of both groups (left with solvent = control, right with NAH) are shown in (B). Miles assay revealed that exogenous Angpt-2 leads to heparanase-dependent eGC breakdown, which contributed to edema formation (C) and plasma leakage of large molecules (D)

## Conclusion

- Breakdown of the endothelial glyocalyx is mediated by Angpt-2
- The Angpt/Tie2 ligand-receptor system is a concurrent gatekeeper of both layers of the vascular double barrier – the endothelial cell and the eGC
- The Tie2 axis might be a future treatment goal to protect the vascular double barrier to prevent vascular leakage in critical care nephrology