INTERNATIO

CONGRESS

Introduction

Targeted gene expression of myelin basic protein (MBP) in hepatocytes can induce MBPspecific CD4 T cell tolerance and prevent MBP-driven neuroinflammation in experimental autoimmune encephalomyelitis (EAE)^{1.} The induction of tolerance depended on IFNG-CXCL9-CXCR3-mediated accumulation of autoreactive T cells in the liver and upregulation of CTLA-4². Alternatively, MBP-specific CD4 T cell tolerance and suppression of EAE can be induced by delivery of MBP peptides to liver sinusoidal endothelial cells (LSECs) via nanoparticles (NP)^{3.}



Aim

It is unclear whether autoantigen delivery to LSECs with NP uses the same or different hepatic tolerance mechanisms as those activated by gene targeting to hepatocytes.



Method

B10.PL mice were immunised to MBP and injected with LSEC-targeting unloaded (NP) or MBPloaded NP (MBP-NP). One day later, CD45.1- MBP-specific tg4 T cells were adoptively transferred into the CD45.1+ recipients. Additionally, mice received blocking IFNG antibody or isotype control. The transferred tg4 T cells were re-analysed ex vivo by flow cytometry on day seven after immunisation. Moreover, the clinical EAE course was monitored.



Conclusions

These results demonstrate that protection from autoimmune disease by autoantigen-delivery to LSECs is dependent on the induction of the IFNG-CXCL9-CXCR3 axis and upregulation of CTLA-4 on autoreactive T cells. Thus, these mechanisms are similar to those induced by hepatic autoantigen gene transfer, suggesting that LSEC-controlled transmigration of autoreactive lymphocytes into the liver parenchyma is essential for NP-mediated tolerance.



References

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CD4 T cell tolerance induced by nanoparticle-mediated autoantigen delivery to liver sinusoidal endothelial cells depends on interferon-gamma

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Results

MBP-specific T cells accumulate in livers of MBP-NP-treated mice



Infiltrating immune cells in the liver were visualised via HE staining (left) on day seven after immunisation. NPCs were isolated and the frequency of transferred, MBP-specific T cells was analysed via flow cytometry (right).

MBP-specific T cell accumulation is dependent on the **IFNG-CXCL9-CXCR3** axis



On day seven after immunisation, production of IFNG was analyzed in MBP-specific T cells after 4h stimulation with PMA/ionomycin (left). Expression of *Cxcl9* (middle) and *Cxcr3* (right) was determined in whole liver tissue.

CTLA-4 is upregulated on MBP-specific T cells in MBP-NP-treated mice







On day seven after immunisation, expression of CTLA-4 was analysed on MBP-specific T cells in the liver via flow cytometry.



MBP-NP-treated mice were immunised to MBP and injected with 300 µg of anti-IFNG or isotype control antibody 2x/week i.p. On day seven after immunisation, the number of MBP-specific T cells was determined in the liver (left). Expression of Cxcl9 was determined in whole liver tissue (middle). CTLA-4 expression was determined on MBP-specific T cells in the liver (right).

In vivo inhibition of IFNG abolished tolerance to MBP in MBP-NPtreated mice and induced clinical EAE



Mice were immunised to MBP and injected with 300 µg of anti-IFNG or isotype control antibody 2x/week i.p. The EAE score was followed for 18 days and the cumulative disease score was calculated.

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In vivo inhibition of IFNG prevented migration of MBP-specific 1 cells into the liver and decreased their expression of CTLA-4







